

STRUCTURE-ACTIVITY RELATIONSHIPS OF A NOVEL CLASS OF SRC SH2 INHIBITORS

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Abstract: The structure-activity relationships (SAR) of a novel class of Src SH2 inhibitors are described. Variation at the pY+1 and pY+3 side chain positions using 2,4- and 2,5-substituted thiazoles and 1,2,4-oxadiazoles as scaffolds resulted in inhibitors that bound as well as the standard tetrapeptide Ac-pYEEI-NH₂. © 1999 Elsevier Science Ltd. All rights reserved.

Introduction: As a part of our efforts toward developing signal transduction inhibitors into therapeutic drugs, we have been interested in applications involving inhibitors of the Src homology 2 (SH2) domain of the tyrosine kinase pp60c-Src, which has been implicated as a potential target 2,3 for therapeutic intervention for both osteoporosis 4-6 and breast cancer. In the preceding communication, we reported on the structure-based design and synthesis of a novel class of Src SH2 inhibitors, represented by the 2,4-substituted thiazole 1b (Figure 1). This class of inhibitors was designed based on structural studies (X-ray, NMR) of the preferred tetrapeptide sequence pTyr-Glu-Glu-Ile (pYEEI) bound to both the Src and Lck SH2 domains. 9-11 The heterocycle ring was incorporated as a replacement scaffold that would appropriately deliver the pY and pY+3 side chains into their respective pockets while gaining a favorable interaction with the hydrophobic surface resulting from Tyr β D5. 11 It was envisioned that the ready availability of enantiopure amino acids could be exploited for the synthesis of nonracemic heterocycles with diverse pY+1 (R1) and pY+3 (R3) side chain substitution. Subsequent structural analysis of thiazole 1b (IC50 = 26 μ M) in both Src and Lck SH2 provided a new frame of reference for analog design. This communication describes our initial results based on thiazole 1b, leading to Src SH2 inhibitors with binding affinities equivalent to that of the standard tetrapeptide, Ac-pYEEI-NH2.

Figure 1

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Chemistry: Recent improvements in the Hantzsch synthesis of nonracemic peptide thiazoles from enantiopure amino acids^{12,13} led to the choice of the 2,4-substituted thiazole as the initial scaffold. Thiazole targets 1a-1d (Figure 1) were prepared from amino acid-derived thioamides and α -bromo ketones as described in the previous communication.^{8,14}

Due to the somewhat limited availability of diverse α-bromo ketones, either from commercial sources or from short synthetic sequences, an effort was initiated toward using heterocycle scaffolds other than the 2,4-substituted thiazole. We have recently reported on the synthesis of nonracemic 2,4,5-substituted peptide thiazoles from two amino acids and one organometallic reagent. A variation in this synthesis, using Gly as the thiazole ring amino acid, provides access to 2,5-substituted thiazoles, the regioisomers of the Hantzsch 2,4-substituted thiazoles. The R³ side chain is derived from an organometallic reagent, thus providing a complementary method toward disubstituted thiazoles. The synthesis of 2,5-substituted thiazoles toward targets 2a-2c is shown in Scheme 1. Boc-Gly-OH was converted into the corresponding Weinreb amide using standard CDI conditions. Addition of the appropriate Grignard reagent 16 provided the protected amino ketones 8a and 8b. Deprotection of carbamates 8a and 8b with 3N HCl gave the corresponding amine salts, which were then coupled with the protected amino acid succinimide derivatives to afford keto amides 9a-9c. Treatment with Lawesson's reagent effected the cyclization giving thiazoles 10a-10c. Deprotection, either by catalytic transfer hydrogenation (10a), 17 or with TFA (10b, 10c), afforded amines 11a-11c. Standard coupling of amines 11a-11c with Ac-Tyr(PO₃Bn₂)-OH followed by deprotection(s) provided 2,5-substituted thiazole targets 2a-2c. 14

Scheme 1. (a) CDI, CH₂Cl₂, 0 °C, 10 min, then DIEA, NH(Me)OMe•HCl, rt, 21 h; (b) R^3 MgBr, THF, 0 °C, 10 min, then rt, 4-7 h; (c) 3N HCl, 1:1 dioxane-EtOAc, rt, 17 h-2.5 d; (d) Et₃N, DME, (Cbz-Abu-OSu for 9a) or (Boc-Trp-OSu for 9b) or (Boc-Glu(Bn)-OSu for 9c), rt, 6-32 h; (e) Lawesson's reagent, THF, 67 °C, 2-3 h; (f) for 11a (and 2c), HCO₂NH₄, 10% Pd-C, MeOH, 64 °C, 2-6 h; (g) for 11b and 11c, TFA, CH₂Cl₂, rt, 1.25 h; 5% NaHCO₃, EtOAc; (h) 11a, 11b or 11c, EDC•HCl, HOBT, DIEA, CH₂Cl₂, DMF, 0 °C to rt; (i) 95:5 TFA-H₂O, rt.

In a continued search for alternative heterocyclic scaffolds, we became aware of a report from the Luthman group 18 describing the synthesis of nonracemic 1,2,4-oxadiazoles, again utilizing amino acids as the source of the R¹ side chains. The R³ side chains are derived from nitriles, many of which are commercially available, after facile conversion into the corresponding amidoximes. Scheme 2 shows the general sequence followed for the preparation of 1,2,4-oxadiazole targets 3a-3f, corresponding to thiazoles 1a-1d and 2a-2c. The nitrile-derived amidoximes 12 were coupled to the appropriate amino acid derivatives 13, giving the intermediate O-acylamidoxime 14. Cyclization of 14 in refluxing pyridine followed by deprotection gave rise to amine 15. Standard coupling of amine 15 with Ac-Tyr(PO₃Bn₂)-OH followed by deprotection(s) provided 1,2,4-oxadiazole targets 3a-3f. 14

Scheme 2. (a) 13 (X = OSu), DME, rt (alternatively, any Boc-amino acid (13, X = OH), CH_2Cl_2 , DMF, EDC•HCl, HOBT, DIEA, rt); (b) pyridine, reflux; (c) TFA, CH_2Cl_2 , rt; (d) 15, EDC•HCl, HOBT, DIEA, CH_2Cl_2 ; (e) 95:5 TFA- CH_2Cl_2 ; (f) LiOH• CH_2Cl_2 ; (THF- CH_2Cl_2) to CH_2Cl_2 to CH_2Cl_2 ; (e) 95:5 TFA- CH_2Cl_2 ; (f) LiOH• CH_2Cl_2 ; (g) 95:5 TFA- CH_2Cl_2 ; (e) 95:5 TFA- CH_2Cl_2 ; (f) LiOH• CH_2Cl_2 ; (g) 95:5 TFA- CH_2Cl_2 ; (g) 95:5 TFA- CH_2Cl_2 ; (h) LiOH• CH_2Cl

Results and Discussion - Comparison of the heterocyclic scaffolds: With synthetic routes using three heterocyclic scaffolds available, attention was turned to the comparison of Src SH2 binding affinity between these scaffolds. During the synthetic development, it became apparent that the 1,2,4-oxadiazole series provided the most versatility toward analog preparation, and resulted in a shorter, more streamlined synthetic pathway. Thus we were particularly interested in whether the 1,2,4-oxadiazole series bound to Src SH2 with similar affinity as the original 2,4-substituted thiazole series (as in 1b). Table 1 contains the assay results ¹⁹ for the selected thiazole and oxadiazole derivatives (1a-3f) prepared for this scaffold comparison. As shown in Table 1, there exists a close correlation in binding affinity between the heterocycle series throughout the range of binding affinities, ²⁰ including the initially-prepared set of 2,4-substituted thiazoles 1a-1d and 1,2,4-oxadiazoles 3a-3d. In addition, the binding affinities for the 2,5-substituted thiazoles, 2a and 2b, were similar to those for the corresponding 1,2,4-oxadiazoles 3e and 3f, which resulted from the subsequent R¹/R³ scan (vide infra). Finally, 2,5-substituted thiazole 2c, prepared for the comparison with both the 2,4-substituted thiazole and 1,2,4-oxadiazole series, had a binding affinity that was similar to those shown for 1b and 3b.²⁰

R ¹	R ³	(2,4- Cmpd	thiazole) IC ₅₀ (μM)		oxadiazole) IC ₅₀ (μM)	` '	thiazole) IC ₅₀ (µM)
Н	CH ₂ Chx	1a	483	3a	896		
CH ₂ CH ₂ CO ₂ H	$CH_2CH(CH_3)_2$	1 c	218	3 c	166		
CH ₂ CH ₂ CO ₂ H	(CH2)2CH(CH3)2	1 d	76	3d	61		
CH ₂ CH ₂ CO ₂ H	CH ₂ Chx	1b	26	3 b	29	2 c	16
CH_2CH_3	(CH2)5CH3			3 e	25	2a	12
CH ₂ -(3-indole)	(CH2)5CH3			3f	16	2 b	14

Table 1: Src SH2 binding affinity comparing similarly substituted thiazoles and oxadiazoles. 19

Results and Discussion - pY+1/pY+3 SAR: With results suggesting that the various heterocycles could be interchanged and a facile 1,2,4-oxadiazole analog synthesis, we began to analyze the SAR for various R^1 and R^3 substitution patterns.²¹ In the tetrapeptide and dipeptide series, it was shown that $pY+1 = Glu (R^1)$ provided inhibitors with the best binding affinity.^{1,22} Table 2 shows the binding affinities for several Gluderived 1,2,4-oxadiazoles, using 3b as the new frame of reference. Shorter branched alkyl chains (3d and 3c) resulted in a loss in binding affinity, as did replacement of the cyclohexyl ring (3b) with a phenyl ring (4d). The n-pentyl analog 4c bound as well as the corresponding CH_2Chx analog 3b. Compounds 4b and 4a, with the slightly longer n-hexyl and n-heptyl side chains, displayed IC_{50} 's that were 3-4 times better than 3b, or equivalent to the standard tetrapeptide sequence Ac-pYEEI-NH₂ ($IC_{50} = 6 \mu M$). Efforts are currently underway toward obtaining additional structural information that may help explain the improved binding affinities for these n-alkyl R^3 analogs.

Table 2: Src SH2 binding affinity for L-Gluderived 1,2,4-oxadiazoles. 19

Cmpd R^3 IC₅₀ (μM) 4a (CH₂)₆CH₃7 4 b (CH₂)₅CH₃ 8 4 c (CH₂)₄CH₃21 3b CH₂Chx 29 3d(CH₂)₂CH(CH₃)₂61 4d CH₂Ph 92 3 c CH₂CH(CH₃)₂ 166

Table 3: Src SH2 binding affinity for L-Glnderived 1,2,4-oxadiazoles. ¹⁹

Cmpd	R ³	IC ₅₀ (μM)		
5a	(CH ₂) ₅ CH ₃	24		
5 b	CH ₂ Ph	72		
5 c	CH ₂ (1-naphthyl)	135		
5d	$(CH_2)_2Ph$	156		
5 e	CH ₂ Chx	278		
5 f	(CH2)2CH(CH3)2	320		
5 g	(CH2)3CH3	384		

With the objective of reducing the overall charge of these molecules in order to improve cellular permeability, part of our R^1/R^3 scan strategy involved replacing the $R^1 = CH_2CH_2CO_2H$ (Glu) side chain with several uncharged R^1 side chains. As was demonstrated in the tetrapeptide and dipeptide series, substitution of the Glu side chain with simple alkyl groups resulted in some loss in binding affinity. 1,22 Our initial choice was that of $R^1 = CH_2CH_2CONH_2$ (Gln), since an early comparison between Gln and Glu derivatives (5b vs. 4d) suggested a reasonable correlation in binding affinities (Tables 2 and 3). Unfortunately, it was soon discovered that Gln was not an acceptable R^1 replacement as demonstrated in the significant decrease in binding affinity upon simple variation (Table 3). Analog 5e was about 10 times less active than the Glu-derivative 3b. The *n*-hexyl analog, 5a, however, was only 3 times less active than the corresponding Glu derivative 4b, again demonstrating that simple *n*-alkyl side chains could be acceptable as R^3 replacements for CH_2Chx .

In our continued efforts toward side chain charge reduction, several analogs in the Trp and Abu series were prepared. As shown in Tables 4 and 5, several analogs (3f, 6a, 7a, 3e) were only 2-3 times less active than the corresponding Glu analogs. This combination of better R¹ and R³ side chains resulted in analogs with binding affinities equivalent to our original lead, 1b, yet with one less charge unit.

As alluded to previously, the 2,5-substituted thiazoles, 2a and 2b (Scheme 1), that corresponded to 3e and 3f, were also prepared. As shown in Table 1, 2a and 2b were found to have binding affinities that were slightly better than 3e and 3f, almost 2 times more active than the original lead 1b, and with one less charge unit.

Table 4: Src SH2 binding affinity for L-Trp-derived 1,2,4-oxadiazoles. 19

 R^3 $IC_{50} (\mu M)$ Cmpd 16 3f (CH₂)₅CH₃ 25 6a CH₂Ph 33 6b (CH₂)₂CH(CH₃)₂35 6 c (CH₂)₆CH₃36 6d (CH₂)₄CH₃62 6e CH₂Chx 89 6f (CH₂)₃CH(CH₃)₂CH₂(1-naphthyl) 279 6 g

Table 5: Src SH2 binding affinity for L-Abuderived 1,2,4-oxadiazoles.¹⁹

Cmpd	R ³	IC ₅₀ (μM)		
7a	(CH ₂) ₆ CH ₃	22		
3 e	(CH2)5CH3	25		
7 b	(CH2)4CH3	65		
7 c	(CH2)2CH(CH3)2	111		

Conclusion: We have described the initial results of variations at R¹, R³ and heterocycle-type based on our initial lead thiazole 1b. The 2,4- and 2,5-disubstituted thiazole and 1,2,4-oxadiazole ring appear to be interchangeable as scaffolds for these disubstituted heterocycle analogs.²⁰ Variation at R³ has resulted in Src SH2 inhibitors (4a, 4b) that bind as well as the standard tetrapeptide Ac-pYEEI-NH2. Variation at R1 has resulted in a reduction of overall charge with only a two fold loss in binding affinity (3f, 2a-2c). Efforts are currently underway at using our 2,4,5-substituted thiazole chemistry¹⁵ toward analogs with improved binding affinity and toward incorporation of pTyr replacements. These results will be reported in due course.

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References and Notes:

- For a recent review on SH2 domains, see: Sawyer, T. K. Biopolymers 1998, 47, 243-261.
- 2. Thomas, S. M.; Brugge, J. S. Annu. Rev. Cell Dev. Biol. 1997, 13, 513-609.
- 3. Brown, M. T.; Cooper, J. A. Biochim. Biophys. Acta 1996, 1287, 121-149.
- 4. Hall, T. J.; Schaeublin, M.; Missbach, M. Biochem. Biophys. Res. Comm. 1994, 199, 1237-1244.
- 5. Boyce, B. F.; Yoneda, T.; Lowe, C.; Soriano, P.; Mundy, G. R. J. Clin. Invest. 1992, 90, 1622-1627.
- 6. Soriano, P.; Montgomery, C.; Geske, R.; Bradley, A. Cell 1991, 64, 693-702.
- Luttrell, D. K.; Lee, A.; Lansing, T. J.; Crosby, R. M.; Jung, K. D.; Willard, D.; Luther, M.; Rodriguez, M.; Berman, J.; Gilmer, T. M. Proc. Natl. Acad. Sci. U.S.A. 1994, 91, 83-87. 7.
- Buchanan, J. L.; Bohacek, R. S.; Luke, G. P.; Hatada, M.: Lu, X.; Dalgarno, D. C.; Narula, S. S.; 8. Yuan, R.; Holt, D. A., Bioorg. Med. Chem. Lett., 1999, 9, 2353.
- 9. Waksman, G.; Shoelson, S. E.; Pant, N.; Cowburn, D.; Kuriyan, J. Cell 1993, 72, 779-790.
- 10. Tong, L.; Warren, T. C.; King, J.; Betageri, R.; Rose, J.; Jakes, S. J. Mol. Biol. 1996, 256, 601-610.
- 11. The protein numbering has been defined by Eck, M. J.; Shoelson, S. E.; Harrison, S. C. Nature 1993, 362, 87-91.
- 12.
- Aguilar, E.; Meyers, A. I. Tetrahedron Lett. 1994, 35, 2473-2476 and references cited therein. Bredenkamp, M. W.; Holzapfel, C. W.; van Zyl, W. J. Synth. Commun. 1990, 20, 2235-2249. 13.
- 14. For representative experimental procedures, see: Buchanan, J. L.; Bohacek, R. S.; Vu, C. B.; Luke, G. P. "Heterocyclic Signal Transduction Inhibitors, Compositions Containing Them & Uses Thereof." PCT US99/05970, March 18, 1999. All compounds were purified to homogeneity by reverse phase HPLC or crystallization and exhibited satisfactory analytical and spectroscopic
- 15. Buchanan, J. L.; Mani, U. N.; Plake, H. R.; Holt, D. A. Tetrahedron Lett. 1999, 40, 3985-3988.
- 16. Nahm, S.; Weinreb, S. M. Tetrahedron Lett. 1981, 22, 3815-3818.
- 17.
- Anwer, M. K.; Spatola, A. F. Synthesis 1980, 929-932.

 Borg, S.; Estenne-Bouhtou, G.; Luthman, K.; Csöregh, I.; Hesselink, W.; Hacksell, U. J. Org. Chem. 18. **1995**, 60, 3112-3120.
- 19. Src SH2 binding affinity was determined using a fluorescence polarization method (Lynch, B. A.; Loiacono, K. A.; Tiong, C. L.; Adams, S. E.; MacNeil, I. A. Anal. Biochem. 1997, 247, 77-82). IC_{50} 's are the average of triplicate measurement. The K_d of the compounds tested in this communication can be estimated by dividing the IC50 by a factor of four.
- 20. This close correlation suggests that either (a) the R¹/R³ substituents in Table 1 continue to align the heterocycle ring away from the Tyr \(\beta D5 \) hydrophobic surface as shown in Figure 3 of ref. 8, or (\beta) the differences in the electronic character of the chosen heterocycles do not significantly affect the subsequent ligand binding affinities.
- 21. The nitriles chosen for pY+3 were based on a biased selection of commercially available nitriles that could potentially bind in the pY+3 pocket. In addition, several nitriles were chosen based on structural analysis based on the X-ray structure of 1b (R. S. Bohacek, unpublished results).
- Gilmer, T.; Rodriguez, M.; Jordan, S.; Crosby, R.; Alligood, K.; Green, M.; Kimery, M.; Wagner, C.; Kinder, D.; Charifson, P.; Hassell, A. M.; Willard, D.; Luther, M.; Rusnak, D.; Sternbach, D. D.; Mehrotra, M.; Peel, M.; Shampine, L.; Davis, R.; Robbins, J.; Patel, I. R.; Kassel, D.; Burkhart, W.; 22. Moyer, M.; Bradshaw, T.; Berman, J. J. Biol. Chem. 1994, 269, 31711-31719.